

## Identification of Cadmium Resistant Bacteria Isolated from Silk Dyed Water. Case Study: Silk Weaving Small Enterprises Ban Chanplane Prokronchai District, Buriram Province

**Tepupsorn Saensuk**

Lecturer, Faculty of Science, Buriram Rajabhat University, Thailand

E-mail: microbiology\_noina@hotmail.co.th

### Abstract

Bioremediation had been shown to be a powerful system for heavy metal pollution cleanup and prevention. Characterization of the cadmium (Cd)-resistant bacteria isolated from silk dyed water were performed in this research. I found that all 14 isolated bacteria have a toxicity tolerant of tolerate toxic CdCl<sub>2</sub> concentrations at 1,500 μM. Interestingly, the Cd binding capacity of all isolated bacteria was very high, ranging from 2.61 to 6.52 log [Cd(atom)]/cell when grown in the presence of 500 μM CdCl<sub>2</sub>. Furthermore, the stability of Cd-bacteria complex of all isolated bacteria was effected by 1 mM EDTA. When grown in the presence of 500 μM CdCl<sub>2</sub>, 3 isolates of Cd-resistant bacteria, including BRU03, BRU07, and BRU11 increasingly produced inorganic sulfide (1X to 3X). These results suggested that these Cd-resistant bacteria have potential ability to precipitate a toxic soluble CdCl<sub>2</sub> as non toxic insoluble CdS.

**Keywords:** Bioremediation, Cadmium resistant bacteria, Cadmium, Silk dyed water

### 1. Introduction

The textile industry is one of the largest polluters in the world. The World Bank estimates that almost 20% of global industrial water pollution comes from the treatment and dyeing of textiles. Some 72 toxic chemicals reach our water supply from textile dyeing. Many of these chemicals cannot be filtered or removed. The textile industry is second only to agriculture as the biggest polluter of clean water globally. Environmental pollution by dyed is a world wide problem causing vast areas of agricultural land to become non-arable and hazardous for animal, plant, and human populations (Eleniet *al.*, 2007). Textile manufacturing dyes release aromatic amines (benzidine and toluidine), heavy metals, ammonia, alkalai salts, toxic solids and large amounts of pigments, chlorine, and a known carcinogen. One of the heavy metals whose concentration is increasing in environments is cadmium (Cd). Cadmium was used in different industrial processes as production of television tube phosphorus, alloy preparation, metal plating, nuclear reactor shields and rods, pigments, stabilizers, batteries, etc. (Hutton *et al.*, 1987; Nriagu, Pacyna, 1988). Cd ion was a highly toxic metal and important element in food-chain contamination. Cadmium, in a variety of chemical forms, is unknown in biological functions (Pinto *et al.*, 2004). Farmers in Asian countries, especially in Japan, confronted with Itai-Itai disease which was resulted from rice grains containing large amount of Cd (Akiko *et al.*, 2006). The conventional physicochemical technologies such as electrochemical treatment, oxidation–reduction, ion exchange and membrane separation were inadequate and expensive to removing metals at lower concentrations (less than 100 mg/l). Bioremediation, which used microbes to detoxify and degrade environmental

contaminants, had received increasing attention in recent times to clean up a polluted environment (Farhadian *et al.*, 2008; Radhika *et al.*, 2006). Since microorganisms had developed survival strategies in heavy metal polluted habitats, their different microbial detoxifying mechanisms such as bioaccumulation, biotransformation, biomineralization and biosorption could be applied either *ex situ* or *in situ* to design economical bioremediation processes (Gadd, 2000; Lim *et al.*, 2003).

Recently, there were reports about resistance, bioaccumulation and biotransformation capacity of Cd by isolated microorganisms under growing conditions such as *Bacillus* sp., *Pseudomonas* sp., *Aspergillus* sp., and *Trichoderma* sp. (Roane, Pepper, 2000; Haq *et al.*, 1999)

## 2. Research Objectives

The objectives of this research were to isolate and identify as well as characterize cadmium-resistant bacteria for use in bioremediation from silk dyed at Silk Weaving Small Enterprises Ban Chanplane Prokronchai District, Buriram Province.

## 3. Research Methodology

### 3.1 Isolation of cadmium-resistant bacteria

Cd-resistant bacteria were isolated from silk dyed water. A portion 25 ml of silk dyed water was suspended in 25 ml nutrient broth (NB) and then incubated at 30 °C on orbital shaker at 150 rpm for 4 h. The 1 ml of suspension was serially diluted (in the range of  $10^{-1}$  to  $10^{-8}$ ) and the each dilution solution was spread on nutrient agar (NA) in the absence and presence of CdCl<sub>2</sub> at 500 µM then incubated at 30 °C for 24 to 48 h.

### 3.2 Cell growth in response to Cd

11 isolates were grown in NB tube for starter bacteria. 1% starter were inoculated into the absence and presence of CdCl<sub>2</sub> at 500 µM and incubated at 30 °C, 150 rpm and take sample for measure at 600 nm every 3 h until 36 h.

### 3.3 Identification of cadmium-resistant bacteria

Cd-resistant isolates were identified by Gram-stain and biochemical conventional tests including conventional tube triple sugar iron agar, lysine iron agar, motility-indole-ornithine decarboxylase agar, phenylalanine-urea broth, β-galactosidase (ONPG), citrate agar, the Voges-Proskauer test and deoxyribonuclease. In addition, the Cd-resistant bacteria were identified by API20NE system. All biochemical tests of all sample were performed in this study.

### 3.4 Cd binding capacity of Cd-resistant isolates

11 isolates were grown in NB medium containing 500 µM CdCl<sub>2</sub> incubated at 30 °C, 150 rpm for 24 h and harvested by centrifugation at 5000x g, 4 °C for 10 minutes. The pellets were then washed twice with sterile water, rewashed twice with EDTA solution (10 ml of 1 mM EDTA in 50 mM HEPES and 10 mM NaCl) and sterile water, respectively. The EDTA treated cells were resuspended with 10 ml sterile water and the Cd content was measured by atomic absorption spectrophotometer.



## 4. Research Results

### Isolation of cadmium-resistant bacteria

During the selection of Cd-resistant bacteria, a total 24 isolates were able to tolerate to toxic Cd at 1500  $\mu\text{M}$ . Only 11 isolates were randomly selected for further study.

### Cell growth in response to Cd

To determine whether the Cd toxicity affected the growth of isolated bacteria, bacteria cells were grown in the absence and presence of toxic concentration of  $\text{CdCl}_2$  at 500  $\mu\text{M}$ . Cell growth was determined by measuring the optical density at 600 nm (Figure 1). In the absence of Cd, all isolated bacteria grew slightly better than in the presence of toxic concentration.

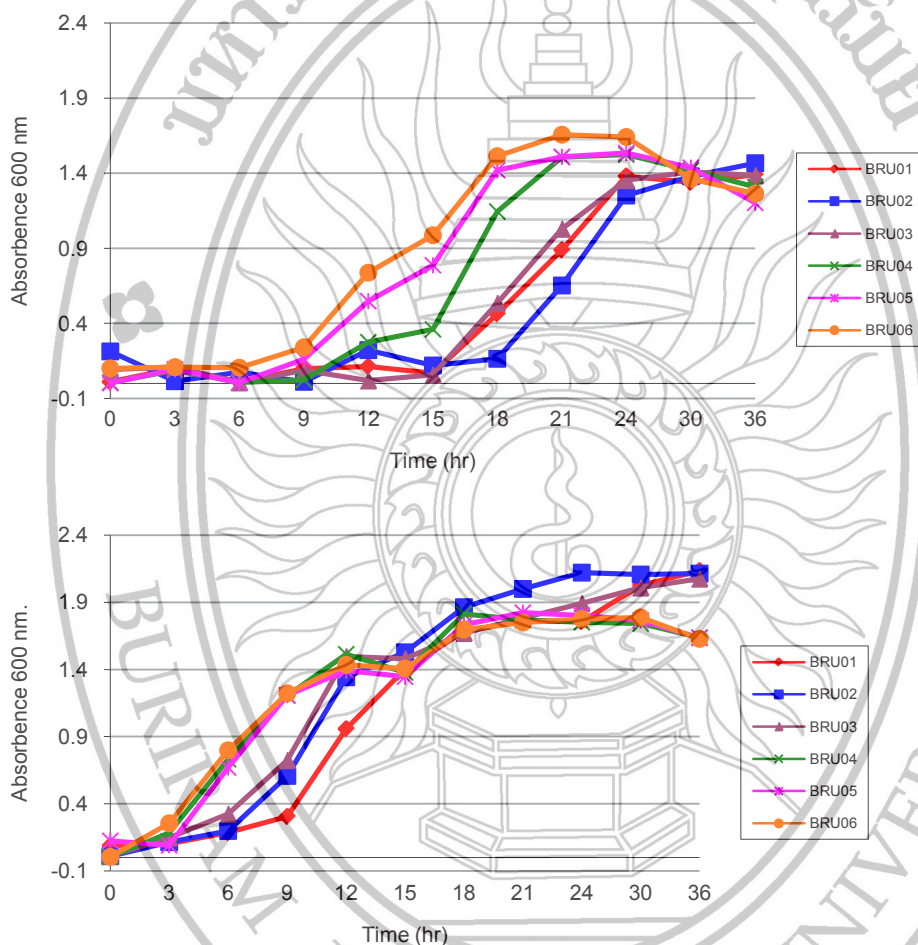


Figure 1. Growth curve of Cd-resistant bacteria in the presence  $\text{CdCl}_2$  (A) and the absence of 500  $\mu\text{M}$   $\text{CdCl}_2$  (B)

### Identification of cadmium-resistant bacteria

11 Cd-resistant isolates were identified to gram-negative bacteria and then biochemical reactions of commercial identification system as follows; Biochemical tests and API20NE. Biochemical tests could one genres. They are *Pseudomonas* sp. All *Pseudomonas* sp were identified their species by used API20NE system. All 11 isolates were *P. aeruginosa* which could be resistant of Cd up to 2500  $\mu\text{M}$ . Structure of *P. aeruginosa* was shown in Table 1.

*Table 1*  
*Phenotypic characteristics of isolates BRU03, BRU07, and BRU11 determined by biochemical tests*

Characteristics	BRU03	BRU07	BRU11
Gram reaction	-	-	-
Cell shape	Rod	Rod	Rod
Cell diameter, $\mu\text{m}$	0.8-1.4	0.8-1.4	0.8-1.4
Oxidase	+	+	+
Catalase	+	+	+
TSI medium	K/N/H <sub>2</sub> S	K/N/H <sub>2</sub> S	K/N/H <sub>2</sub> S
Citrate	+	+	+
Motility	+	+	+
Mac	NFL	NFL	NFL
MR-VP	-/-	-/-	-/-
Growth at 4 °C	-	-	-
Lysine iron agar	-	-	-
$\beta$ -galactosidase	-	-	-
Urea	-	-	-
Starch	-	-	-
Gelatin	-	-	-
6% NaCl	-	-	-
Indole	-	-	-

#### **Cd binding capacity of Cd-resistant isolates**

In order to determine the ability of Cd-resistant bacteria to accumulate Cd from the environment, the Cd binding capacity of 24 isolates was measured. As show in Table 1, the Cd binding capacity of all isolated bacterial cells was 5.42 to 6.52  $\log[\text{Cd}(\text{atom})/\text{cell}]$ . I further studied the effect of EDTA on their Cd binding capacity

(Table 2). The results showed that the Cd binding capacity of all isolates were affect by 1 mM EDTA.

Table 2

*Cd binding capacity of Cd-resistant bacteria treated and untreated with EDTA*

Isolate	Cd binding capacity	
	Log Cd(atom)/Cell	
	[No EDTA]	[EDTA]
BRU01	6.58±0.18	5.20±0.14
BRU02	5.30±0.27	6.21±0.52
BRU03	4.38±0.13	6.52±0.32
BRU04	5.76±1.02	3.24±0.43
BRU05	5.39±0.86	3.73±0.61
BRU06	3.38±0.73	4.40±0.42
BRU07	4.90±0.54	5.52±0.84
BRU08	4.51±0.61	2.61±0.17
BRU09	4.96±0.72	4.83±0.95
BRU10	4.50±0.59	4.96±0.82
BRU11	4.66±0.77	5.20±0.84

## 5. Discussion

Agricultural soils were primarily contaminated with Cd<sup>2+</sup> due to the excessive use of phosphate fertilizers, dispersal of sewage sludge and atmospheric deposition. Cd<sup>2+</sup> was readily taken up by numerous crops including cereals, potatoes, rice and fruits (Ingwersen and Streck, 2005). Consumption of rice grown in paddy soils contaminated with Cd<sup>2+</sup> may pose a serious risk to human health, because 22–24% of the total metal content in rice biomass was concentrated in the rice grains (Wang et al., 2003). Thus, contamination by Cd<sup>2+</sup> is increasing in both human food and overall in the agricultural environment. Plants readily takes up Cd<sup>2+</sup> from the soil. However, exposure to high levels of Cd<sup>2+</sup> resulted in reduced rates of photosynthesis, chlorosis,



growth inhibition, browning of root tips, decreased water and nutrient uptake, and ultimately death (Marcano et al., 2002). Even metals exert their toxic effects on microorganisms through various mechanisms, and metal-tolerant bacteria could survive in these habitats and could be isolated and selected for their potential application in the bioremediation of contaminated sites (Piotrowska-Seget et al., 2005).

Many microorganisms in the soil are able to solubilize “unavailable” forms of heavy metal-bearing minerals by excreting organic acids. And many soil bacteria are tolerant to heavy metals and play important roles in mobilization of heavy metals (Gadd, 1990). In this study were found 11 bacterial isolates which could be resistant to Cd. 11 isolates were *P. aeruginosa*. In this results indicated that the isolated BRU03 may be useful in preventing Cd diffusion in the soil environment. Further investigation of molecular characterization of Cd-resistant bacteria.

#### Acknowledgements

I wish to acknowledge the Faculty of Science, KhonKaen University and The research and development institute of BuriramRajabhat University for permission to use scientific instruments during this study.

#### References

- Akiko Ike, Rutchadaporn Sriprang, Hisayo Ono, Yoshikatsu Murooka, and Mitsuo Yamashita, 2006. Bioremediation of cadmium contaminated soil using symbiosis between leguminous plant and recombinant rhizobia with the MTL4 and the PCS genes. *Chemosphere* 66, 1670–1676.
- Belimov, A.A., Safronova, V.I., Sergeyeva, T.A., Egorova, T.N., Matveyeva, V.A., Tsyganov, V.E., Borisov, A.Y., Tikhonovich, I.A., Kluge, C., Preisfeld, A., Dietz, K.J., and Stepanok, V.V. 2001. Characterization of plant growth-promoting rhizobacteria isolated from polluted soils and containing 1-aminocyclopropane-1-carboxylate deaminase. *Can. J. Microbiol.* 47, 642–652.
- Farhadian, M., Vachelard, C., Duchez, D., Larroche, C., 2008. In situ bioremediation of monoaromatic pollutants in groundwater: A review. *Bioresour. Technol.* 99, 5296–5308.
- Gadd, G.M., 1990. Heavy metal accumulation by bacteria and other microorganisms. *Experientia* 46, 834–840.
- Gadd, G.M. 2000. Bioremediation potential of microbial mechanisms of metal mobilization and immobilization. *Curr. Opin. Biotechnol.* 11, 271–279.
- Harry, Aiking., Karin, Kok., Harm Van, Heerikhuizen., and Jan, Van’T Riet., 1982. Adaptation to Cadmium by *Klebsiella aerogenes* Growing in Continuous Culture Proceeds Mainly via Formation of Cadmium Sulfide. *Appl. Environ. Microb.* 44, 938–944.
- Haq R, Zaidi SK, Shakoori AR. 1999. Cadmium resistant *Enterobacter cloacae* and *Klebsiella* sp. isolated from industrial effluents and their possible role in cadmium detoxification. *World J Microbiol Biotechnol.* 15, 283–90.
- Hutton, M., Chaney, M., Krishna Murti, C.R., Olade, A., Page, A.L., 1987. Group report: cadmium. In: Hutchinson, T.C., Meema, K.M. (Eds.), *Lead, Mercury, Cadmium and Arsenic in the Environment*. John Wiley & Sons, New York, 35–41.

- Ingwersen, J., Streck, T., 2005. A regional-scale study on the crop uptake of cadmium from sandy soils: measurement and modeling. *J. Environ. Qual.* 34, 1026–1035.
- Marcano, L., Carruyo, I., Del Campo, A., Montiel, X., 2002. Effect of cadmium on the nucleoli of meristematic cells of onion *Allium cepa*L.: an ultrastructural study. *Environ. Res.* 88, 30–35.
- Nriagu, J.O., Pacyna, J.M. 1988. Quantitative assessment of worldwide contamination of air, water and soils by trace metals. *Nature* 333, 134–139.
- Pinto AP, Mota AM, de Varennes A, Pinto FC. Influence of organic matter on the uptake of cadmium, zinc, copper and iron by sorghum plants. *Sci Total Environ* 2004;326:239–47.
- Piotrowska-Seget, Z., Cycon, M., Kozdroj, J., 2005. Metal-tolerant bacteria occurring in heavily polluted soil and mine spoil. *Appl. Soil Ecol.* 28, 237–246.
- Radhika, V., Subramanian, S., and Natarajan, K.A. 2006. Bioremediation of zinc using *Desulfotomaculum nigrificans*: bioprecipitation and characterization studies. *Water Res.* 40, 3628–3636.
- Roane TM, Pepper IL. 2000. Microbial responses to environmentally toxic cadmium. *Microb Ecol.* 38, 358–64.
- Wang, C.L., Michels, P.C., Dawson, S.C., Kitisakkul, S., Baross, J.A., Keasling, J.D., and Clark, D.S., 1997. Cadmium removal by a new strain of *Pseudomonas aeruginosa* in aerobic culture. *Appl. Environ. Microb.* 63, 4075–4078.
- Wang, Q.R., Cui, Y.S., Liu, X.M., Dong, Y.T., Christie, P., 2003. Soil contamination and plant uptake of heavy metals at polluted sites in China. *J. Environ. Sci. Health Part A: Toxic Hazard Subst. Environ. Eng.* 38, 823–838.
- White, C., Gadd, G.M., 1996. Mixed sulphate-reducing bacterial cultures for bioprecipitation of toxic metals: factorial and response-surface analysis of the effects of dilution rate, sulphate and substrate concentration. *Microbiology* 142, 2197–2205.